APC-IgG Conjugation Kit For labeling 3 x 10 μg IgG

Reagent Storage

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The kit is shipped on blue ice. Please store kit components as described below

Kit Component	Storage Temp	Storage Notes	
Concentrated Activator	-20°C	Keep the vial in the desiccated container as supplied in the kit	
APC-Z™	-20°C or 2-8°C	Does not need to be kept desiccated.	
Quenching Reagent	-20°C or 2-8°C	Does not need to be kept desiccated.	
Zeba Desalting Columns with Collection Tubes	2-8°C	Does not need to be kept desiccated. Do not store at freezing temperatures	

Introduction

Allophycocyanin is widely used as a fluorescent label in immunochemistry techniques such as flow cytometry and cellular analysis. Preparation of bright, stable, and reproducible antibody-APC conjugates is one of the biggest challenges for development of high-quality fluorescent reagents for cellular analysis and flow cytometry. The Moss APC-IgG conjugation kit utilizes a novel chemistry to generate bright and highly reproducible APC-IgG conjugates with a simple procedure

Features

- Liquid-based reagents.
- Completely scalable: conjugate anywhere from 10 µg to 1-gram IgG per reaction.
- Supplies sufficient activated APC to conjugate all IgG at a 2:1 APC:IgG ratio.
- Highly efficient APC incorporation purification not usually necessary.
- Customize the APC:IgG ratio to create optimized conjugates for different applications.
- Conjugates have greatly improved stability vs Lightning-Link™ and traditional chemistry.

Product & Contents

Catalogue Number	APC-Link-CA	
For Labeling:	3 x 10 μg IgG	
Concentrated Activator	10 µL	
APC-Z™ - Activated APC (7 mg/ml)	12 µL	
1X Quenching Reagent	25 μL	
Zeba Desalting Column with Collection Tube	3 each	



Additional Reagents Required But Not Supplied

1X Phosphate Buffered Saline (1X PBS), pH 7.2-7.5 Deionized water (dH₂O) 1.5 ml microcentrifuge tubes

Shelf Life

The performance of the product is guaranteed for a minimum of 12 months when stored as directed.

IgG Requirements

The IgG to be labeled should be at a minimum concentration of 0.8 mg/ml in pure 1X PBS and should not contain any preservatives or carriers such as sodium azide, Proclin 300 or BSA.

APC:IgG Molar Ratio

This kit utilizes a 2:1 APC:IgG molar ratio which is optimal for most conjugations reaction. However, lower or higher ratios may give better results depending upon the antibody characteristics and the intended end-use. To change the APC:IgG molar ratio, vary the volume of APC-Z[™] added to the conjugation reaction (Step 8).

Conjugation Procedure - Overview





Before Beginning The Procedure

Remove the Concentrated Activator from the freezer. <u>Important: Allow sufficient</u> <u>time to let the container and contents come to room temperature before opening</u> <u>the outer and inner vials.</u>

Note: The jar containing the Activator can be removed from the freezer up to 24 hours before use.

Detailed Conjugation Procedure

- 1. Measure the absorbance of the IgG solution at 280 nm using PBS as a blank. Divide the A280 by 1.40 to obtain the IgG concentration in mg/ml.
- 2. Dilute IgG to 1.20 mg/ml in 1X PBS (0.8 1.4 mg/ml is acceptable).
- 3. Add 10 μL of IgG solution to a new microcentrifuge tube.
- 4. Prepare a <u>working dilution (1X)</u> of Activator from Concentrated Activator in deionized water:
 - a. Add 2.0 μL of Concentrated Activator to 1300 μL of deionized water (1:650 dilution).
 - b. Immediately vortex to mix the solution thoroughly.
- Note: The <u>1X</u> Activator must be used within 5 minutes of preparation. If more than 5 minutes passes before use, discard the 1X Activator and prepare a fresh solution.
- 5. Add 2.0 μL of <u>1X</u> Activator to the 10 μL aliquot of IgG and then mix thoroughly by gentle vortexing.
- 6. Incubate the solution at room temperature for 1 hour.

Note: A longer incubation is not harmful and even overnight incubations will be successful.

7. Desalt the complete 12 μ L reaction volume into pure 1X PBS using the included Zeba spin column. See the attached desalting protocol.

Note: The activated IgG is stable and can be stored at 2-8°C for at least 4 months.

- 8. Add 2 μ L of APC-ZTM to the desalted, activated IgG and mix by gentle vortexing.
- 9. Incubate the solution at room temperature for 2-24 hours.



Note: Usable conjugates are produced after only 2 hours incubation. Larger and more potent conjugates will be produced after longer incubations.

10. Add 2 μ L of Quenching Reagent to the reaction and mix by gentle vortexing.

11. Incubate the solution at room temperature for 1 hour.

Note: A longer incubation is not harmful and overnight incubations will be successful.

12. Conjugate is ready for use. Store at 2-8°C.

Note: To improve conjugate performance, it may help to purify the conjugate from the unincorporated APC and reaction components by size exclusion chromatography.

Optional Accessories

For desalting IgG before activation - Order from Thermo Fisher Scientific:

Sample Size	Description	Cat #
2 – 12 µL	Zeba Spin Desalting Columns, Micro (75µL), 7K MWCO	89877, 89878
30 - 130 µL	Zeba Spin Desalting Columns, 0.5mL, 7K MWCO	89882, 89883

For concentrating IgG before or after IgG activation or for concentrating the final conjugate – Order from MilliporeSigma:

Sample Size	Description	Cat #
Up to 500 µL	Amicon Ultra-0.5 Centrifugal Filter Unit with Ultracel-50 membrane	Z740176

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Zeba Spin Desalting Column Instructions

75 μ**L columns for 2 – 12**μ**L Samples**

Notes:

- Each column can desalt a 2-12 μL sample.
- The resin slurry contains 0.03% sodium azide.
- These columns are recommended for desalting molecules > 7000 Daltons.

Storage:

- Store columns at 2-8°C.
- Columns may be stored at room temperature for several days without adverse effects.

Additional Materials Required

- Variable-speed bench-top microcentrifuge
- 1.5 mL microcentrifuge collection tubes

Spin Column Preparation

- 1. Remove column's bottom closure and loosen cap (do not remove cap).
- 2. Place column in the collection tube provided.
- 3. Centrifuge at $1000 \times g$ for 1 minute to remove storage solution.
- 4. Discard storage solution from the collection tube. Remove column cap. It is not necessary to replace the cap for subsequent steps.
- 5. Add 50 μ L of buffer on top of the resin bed. Centrifuge at 1000 × g for 1 minute to remove buffer. Discard the buffer from the collection tube.
- 6. Repeat Steps 5 two additional times for a total of 3 column washes.

Sample Desalting

- 7. Place column in a new microcentrifuge tube and slowly apply the sample to the center of the compacted resin bed.
- 8. Do not add a stacker after application of the sample to the resin bed.
- 9. Centrifuge column at 1000 × g for 2 minutes to collect desalted sample. Discard desalting column after use.

Zeba Desalting Columns are a product of Thermo Fisher Scientific Inc.

Sample Size	Description	Cat #
2 – 12 µL	Zeba Spin Desalting Columns, Micro (75µL), 7K MWCO	89877, 89878